Procarta[™] Transcription Factor Whole Cell Lysis Kit

User Manual



Panomics, Inc.

Procarta Transcription Factor Whole Cell Lysis Kit User Manual

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About the User Manual

this Manual

Who Should Read Anyone that has purchased a Procarta Whole Cell Lysis Kit from Panomics to prepare whole cell lysates for use in Panomics' Procarta Transcription Factor (TF) Assay Kits.

Covers

What this Manual This manual provides the following:

- Kit contents
- Required materials and equipment
- Whole cell lysis procedure

Safety Warnings and Precautions

All chemicals should be considered potentially hazardous. We recommend that CAUTION this product and its components be handled by those trained in laboratory techniques and be used according to the principles of good laboratory practice.

Note This product is intended for research use only.

For More For information about the Procarta products mentioned in this manual, visit our Information website at www.panomics.com.

Procarta Transcription Factor Whole Cell Lysis Assay Kit

Transcription **Factor Whole Cell** Lysis Kit

About the The Procarta TF Whole Cell Lysis Kit contains reagents and procedures for the preparation of whole cell lysates for use in our Procarta TF Assay Kits. The Procarta TF Whole Cell Lysis Kit contains sufficient reagents for the preparation of 40 whole cell lysates from cultured cells grown in 6-well culture plates or 240 whole cell lysates from cultured cells grown in 96-well culture plates.

Kit Contents and The Procarta Transcription Factor Whole Cell Lysis Assay Kit contains the following Storage components.

Procarta TF Whole Cell Lysis Kit components:

Component	Quantity	Storage
Lysis Buffer I	12.0 mL	−20 °C
Lysis Buffer II	1.2 mL	−20 °C
DTT	120 µL	−20 °C
Protease inhibitor cocktail	120 µL	−20 °C

Required Materials and Equipment Not Provided

Materials and Equipment

Item	Source
1X PBS	Invitrogen (P/N 14190-144)
Rocking platform	VWR, Rocking Platform, Model #100 or equivalent
Centrifuge	Eppendorf #5804R
Protein determination kit	Bio-Rad DC Protein Assay Kit (P/N 500-0112) or equivalent
Microcentrifuge tubes	Major laboratory supplier (MLS)
15 mL conical centrifuge tubes	MLS
Adjustable single and multi-channel precision pipettes	MLS
PCR plates	MLS

Cell Preparation

Growing Cells In all cases, cells are grown to about 90% confluence. The following table provides recommendations for the cell requirements for each culture vessel type. However, it is important to realize that cell types vary in size and actual numbers of cells/vessel may vary.

Use the table below as a guide.

Culture Vessel	Cell Number
100-mm culture dish	1 x 10 ⁷ cells/dish
6-well plate	1-5 x 10 ⁶ cells/well
24-well plate	2-5 x 10 ⁵ cells/well
96-well plate	2-5 x 10 ⁴ cells/well

Whole Cell Lysis Procedure

Assay Guidelines IMPORTANT All components and PBS must be kept on ice at all times. Lysis Buffer 1 Working Reagent must be kept on ice and should be used within 2 hours of preparation.

Preparing Working To prepare working reagent: Reagent

Step	Action		
1	For preparation of 1 mL Lysis reagents, then invert tube to n	Buffer 1 Working Reagent, combine the foll nix:	owing
	1 mL Lysis Buffer 1		
	10 μL DTT		
	10 μL Protease Inhibitor Cocktail		
	below as a guide.	according to experimental design. Use the Quantity of Lysis Working	table
	Vessel	Reagent	
		110090111	
	100 mm culture dish	1 mL/dish	
	100 mm culture dish 6-well plate		
		1 mL/dish	

Preparing Whole Cell Lysates From Adherent Cells

Preparing Whole To prepare working reagent:

	Action			
1	Remove the culture media from all wells and wash cells twice with an appropriate volume of cold 1X PBS.			
	Vessel		Quantity of PBS	
	100 mm culture dish		10 mL/dish	
	6-well plate		1 mL/well	
	24-well plate		500 μL/well	
	96-well plate		200 μL/well	
2	Following the second was	h, make s	ure the PBS is completely removed.	
3	Add the appropriate volum	ne of Lysis	Buffer I Working Reagent to the wells.	
	Vessel	Quan	tity of Lysis Buffer I Working Reagent	
	100 mm culture dish	Ţ Ţ		
	6-well plate	250 μ	_/well	
	24-well plate	100 µL/well		
	96-well plate	50 μL/well		
4	Transfer culture vessel(s) to an ice bucket and transfer ice bucket to a rocking platform at 200 rpm for 10 minutes. Add the appropriate volume of Lysis Buffer II Working Reagent to the wells.			
5				
5		ne of Lysis	Buffer II Working Reagent to the wells.	
5	Add the appropriate volum	ne of Lysis	Buffer II Working Reagent to the wells.	
5	Add the appropriate volun	Quan	Buffer II Working Reagent to the wells. tity of Lysis Buffer II Working Reagent /dish	
5	Add the appropriate volun Vessel 100 mm culture dish	Quan	Buffer II Working Reagent to the wells. Lity of Lysis Buffer II Working Reagent _/dish well	
5	Add the appropriate volun Vessel 100 mm culture dish 6-well plate	Quan 100 µ 25 µL	Buffer II Working Reagent to the wells. Lity of Lysis Buffer II Working Reagent Joish Well Well	
5	Vessel 100 mm culture dish 6-well plate 24-well plate 96-well plate	Quan 100 µ 25 µL 10 µL 5 µL/v o an ice b	Buffer II Working Reagent to the wells. Lity of Lysis Buffer II Working Reagent Joish Well Well	
	Vessel 100 mm culture dish 6-well plate 24-well plate 96-well plate Transfer culture vessel(s) t platform at 200 rpm for 1 le	Quan 100 µ 25 µL 10 µL 5 µL/v o an ice b hour. Il times, the	Buffer II Working Reagent to the wells. Lity of Lysis Buffer II Working Reagent L/dish Well Well Well Ucket and transfer ice bucket to a rocking en transfer each sample to a 1.5 mL at 14,000 x g for 3 minutes at 4°C. Note th	ne
6	Vessel 100 mm culture dish 6-well plate 24-well plate 96-well plate Transfer culture vessel(s) t platform at 200 rpm for 1 l Pipet up and down severa microcentrifuge tube and orientation of tube in centre	Quan 100 µ 25 µL 10 µL 5 µL/v o an ice b hour. Il times, the centrifuge as p p and down	Buffer II Working Reagent to the wells. Lity of Lysis Buffer II Working Reagent L/dish Well Well Well Ucket and transfer ice bucket to a rocking en transfer each sample to a 1.5 mL at 14,000 x g for 3 minutes at 4°C. Note th	s

To prepare working reagent: (continued)

Action			
Measure the protein concentration of each sample using a protein quantitation assay (sold separately). Then prepare 5 µL aliquots for each of the samples. Store samples at –80 °C or use immediately in Procarta TF Assay Kit.			
Vessel	Typical Protein Yields		
100 mm culture dish	300–800 μg/dish		
6-well plate	200–500 μg/well		
24-well plate	40–80 μg/well		
96-well plate 20–40 µg/well			
	Measure the protein conceassay (sold separately). The samples at -80 °C or use in the samples		

Preparing Whole Cell Lysates From Suspension Cells

Preparing Whole To prepare whole cell lysates from suspension cells:

500 x g for 5 minutes. For 96-well plates, transfer of minutes Remove the culture media ar followed by centrifugation at For 96-well plates, remove th 200 μL of cold 1X PBS follow wash step. Following the second wash stemoved from the cells.	15 mL centrifuge tube as appropriate and centrifuge at cells to a PCR plate and centrifuge at 500 x g for 5 and wash cells by resuspending in 1 mL of cold 1X PBS to 500 x g for 5 minutes. Repeat wash step. The culture media and wash cells by resuspending in wed by centrifugation at 500 x g for 5 minutes. Repeat step, ensure that the 1X PBS solution is completely centrifugation, transfer contents from the 15 mL	
minutes Remove the culture media ar followed by centrifugation at For 96-well plates, remove th 200 µL of cold 1X PBS follow wash step. Following the second wash stemoved from the cells.	nd wash cells by resuspending in 1 mL of cold 1X PBS t 500 x g for 5 minutes. Repeat wash step. the culture media and wash cells by resuspending in wed by centrifugation at 500 x g for 5 minutes. Repeat step, ensure that the 1X PBS solution is completely entrifugation, transfer contents from the 15 mL	
followed by centrifugation at For 96-well plates, remove th 200 µL of cold 1X PBS follow wash step. Following the second wash stemoved from the cells. Note Before the second ce	t 500 x g for 5 minutes. Repeat wash step. the culture media and wash cells by resuspending in wed by centrifugation at 500 x g for 5 minutes. Repeatestep, ensure that the 1X PBS solution is completely entrifugation, transfer contents from the 15 mL	
200 µL of cold 1X PBS follow wash step. Following the second wash stemoved from the cells. Note Before the second cells.	wed by centrifugation at 500 x g for 5 minutes. Repeats step, ensure that the 1X PBS solution is completely entrifugation, transfer contents from the 15 mL	
removed from the cells. Note Before the second ce	entrifugation, transfer contents from the 15 mL	
	microcentrituge tube.	
Immediately add the appropriate volume of Lysis Buffer 1 Working Reagent to pellets. Mix by pipetting up and down several times.		
Original Culture Vessel	Lysis Buffer I Working Reagent	
100 mm culture dish	1 mL/dish	
6-well plate	250 μL/well	
24-well plate	100 μL/well	
96-well plate	50 μL/well	
	Original Culture Vessel 100 mm culture dish 6-well plate 24-well plate	

To prepare whole cell lysates from suspension cells: (continued)

Step	Action		
5	Add the appropriate volume of Lysis Buffer II to the cells.		
	Original Culture Vessel	Quantity of Lysis Buffer II Working Reagent	
	100 mm culture dish	100 μL/dish	
	6-well plate	25 μL/well	
	24-well plate	10 μL/well	
	96-well plate	5 μL/well	
6	Transfer tube(s) or culture plate to an ice bucket and place on a rocking platform at 200 rpm for 1 hour.		
7	For microcentrifuge tubes, pipet up and down several times and centrifuge at maximal speed (12,000 x g) for 3 minutes at 4°C. Note the orientation of tube in centrifuge as pellets may not be visible.		
	For PCR plates, pipet up and down several times and centrifuge the PCR plate at 2,250 x g for 5 minutes.		
8	Transfer supernatant(s) to a new microcentrifuge tube or a new PCR plate, this is your whole cell lysate.		
9	Measure the protein concentration of each sample using a protein quantitation assay (sold separately). Then prepare 5 µL aliquots for each of the samples. Store samples at –80 °C or use immediately in Procarta TF Assay Kit.		
	Vessel	Typical Protein Yield	
	100 mm culture dish	300–800 μg/dish	
	6-well plate	200–500 μg/well	
	24-well plate	40–80 μg/well	
	96-well plate	20–40 μg/well	

Contacting Panomics

Technical Help For technical questions, contact our technical support group by telephone at 1-877-726-6642 option 3 or by email at techsupport@panomics.com (US and Canada) or techsupport_europe@panomics.com (Europe), or visit our website www.panomics.com for an updated list of FAQs and product support literature.

For Additional For information about Panomics products or for ordering information, contact your **Services** Regional Sales Manager, or visit our website at www.panomics.com.